

International Journal of Biomedicine 14(1) (2024) 93-98 http://dx.doi.org/10.21103/Article14(1)_OA14

ORIGINAL ARTICLE

Andrology



Assessment of Sperm Morphometry in Evaluating Male Infertility

Ramadan S. Hussein^{1*}, Essam-Eldeen M. Mohamed², Refaat R. Mohamed², Walid Kamal Abdelbasset³, Wessam Ezzat Morsy^{4,5}, Shereen H. Elsayed⁶

¹Department of Dermatology, College of Medicine Prince Sattam Bin Abdulaziz University, Al-Kharj, Saudi Arabia ²Department of Dermatology, Andrology and STDs, College of Medicine, Al-Azhar University, Assiut, Egypt ³Department Physiotherapy, College of Health Sciences, University of Sharjah, Sharjah, United Arab Emirates ⁴Department of Physiology, College of Medicine, Ain-Shams University, Cairo, Egypt ⁵Department of Physiology, College of Medicine, Armed Forces College of Medicine, Cairo, Egypt ⁶Department of Rehabilitation Sciences, Faculty of Health and Rehabilitation Sciences, Princess Nourah bint Abdulrahman University, Riyadh, Saudi Arabia

Abstract

Background: Infertility is a complex issue affecting 15% of couples of reproductive age, with men accounting for 40%-50% of infertility cases. Semen analysis comprises various descriptive measures of sperm and seminal fluid to determine semen quality. Transforming qualitative descriptions of sperm deformities and shape changes into quantitative terms can aid in identifying sub-visual abnormalities. This study aimed to evaluate sperm morphometry parameters in both infertile and fertile men.

Methods and Results: The study enrolled a total of 101 participants, divided into three groups: Group A included 38 subfertile patients with varicocele, Group B included 33 patients with idiopathic infertility (23 with asthenozoospermia and 10 with oligozoospermia), and Group C (the control group) included 30 healthy fertile men. The mean age of patients was 31.6 ± 5.81 , 31.3 ± 6.0 , and 29.47 ± 4.27 years in Groups A, B, and C, respectively (*P*>0.05). Scrotal duplex examinations were performed to identify the presence of varicocele. Semen samples were collected following WHO Manual (2010). Semen dynamic and morphological analyses were conducted using CASA (Computer-Assisted Semen Analysis, MIRALAB, ISO9001, ISO13485). We found that sperm concentration, total sperm count, sperm progressive motility, and sperm progressive+non-progressive motility were significantly lower in Group A and Group B than in Group C (*P*=0.000 in all cases); however, there were no differences between Group A and Group B regarding these parameters. The sperm morphology index was significantly lower in Group A than in Group C (*P*=0.0024); no differences were found between Group B and Group C and Group B and Group B and Group C (*P*=0.004).

Conclusion: Our study highlights the significant association between sperm morphology and male infertility in varicocele and idiopathic subfertile males.(International Journal of Biomedicine. 2024;14(1):93-98.)

Keywords: infertility • semen quality • varicocele

For citation: Hussein RS, Mohamed EEM, Mohamed RR, Abdelbasset WK, Morsy WE, Elsayed SH. Assessment of Sperm Morphometry in Evaluating Male Infertility. International Journal of Biomedicine. 2024;14(1):93-98. doi:10.21103/Article14(1)_OA14

Abbreviations

MAI, multiple anomalies index; TZI, teratozoospermia index; SDI, sperm deformity index.

Introduction

Infertility is a condition of the reproductive system characterized by the inability to achieve a clinical pregnancy after 12 months or more of regular, unprotected sexual activity. Infertility is a complex issue affecting 15% of couples of reproductive age,⁽¹⁾ with men accounting for 40%-50% of infertility cases.^(2,3) Various factors, including occupational hazards, exposure to reproductive toxicants, chemotherapy, radiation therapy, heat exposure, physical labor, lifestyle variables (wearing tight underwear, poor diet), genital injuries, hereditary traits, testicular maldescent, infections, and iatrogenic causes, can contribute to decreased male fertility.⁽⁴⁻⁶⁾

The most prevalent form of male infertility is idiopathic male infertility, which is characterized by the presence of one or more abnormal semen parameters without a clear explanation.⁽⁷⁾ Following closely is varicocele, accounting for 35% to 50% of men with primary infertility and up to 81% with secondary infertility.⁽⁸⁾ The negative impact of varicocele on spermatogenesis can be attributed to several factors, including elevated testicular temperature, increased intratesticular pressure, hypoxia due to reduced blood supply, reflux of toxic compounds from the adrenal glands, and hormonal profile abnormalities.^(9,10)

Semenanalysis involves a set of descriptive measurements of spermatozoa and seminal fluid parameters used to assess semen quality.⁽¹¹⁾ Determining sperm morphology, however, poses challenges due to subjective factors and inconsistency. A comprehensive assessment of sperm shape necessitates an evaluation of the head, neck, midsection, and tail. In normal sperm, the head should be oval and symmetrical, and tail insertion should be axial, in line with the long axis of the head. Abnormal sperm variations include those with oversized, undersized, round, asymmetrical, or amorphous heads, as well as those with tapering, bulging midpieces, multiple heads or tails, or amorphous heads.⁽¹²⁾ Typically, clinical laboratories apply sperm morphology parameters established by the WHO or the "strict morphology" criteria developed by Dr. Kruger.⁽¹³⁾

To enhance the quantitative identification of sperm shape, morphometric methods to measure sperm under normal conditions can establish a reference for quantitative terms, replacing qualitative descriptions with more precise numerical terms. Quantitative stereological methods allow investigators of seminal samples to derive three-dimensional concepts from two-dimensional microscopic fields. This enriches the biophysical assessment of sperm by calculating absolute and relative volumetric parameters, which conventional microscopic assessment cannot provide.(14,15) Converting qualitative descriptions of sperm deformities and shape changes into quantitative numerical terms can be particularly valuable in identifying sub-visual shape changes and abnormalities.^(16,17) Using quantitative numerical descriptions for qualitative characteristics can facilitate the comparison of different treatment modalities and determine their respective advantages. Mathematical descriptions of sperm movement allow for a more precise expression of the type of movement and velocity, which can be challenging to convey using ambiguous qualitative terms.(18,19)

A recent study by Rrumbullaku et al.⁽²⁰⁾ demonstrated a significant increase in the percentage of tapered spermatozoa, spermatozoa containing cytoplasmic droplets, and spermatozoa with bent tails in varicocele patients, compared to controls. In our study, we evaluated sperm morphometry in varicocele, non-varicocele infertile patients, and controls using Computer-Assisted Semen Analysis (CASA, MIRA LAB, ISO9001, ISO13485). This approach promises to provide a more accurate and quantitative assessment of sperm morphology, shedding light on potential sub-visual abnormalities and shape changes that could be contributing to male infertility.

This study aimed to evaluate sperm morphometry parameters in both infertile and fertile men.

Materials and Methods

Study Setting and Participants

This prospective study took place at the Andrology Unit of Alazhar University Hospital (Assiut) and was conducted with the approval of the relevant authorities. Informed consent was obtained from all participants. The study enrolled a total of 101 participants, divided into three groups: Group A included 38 subfertile patients with varicocele, Group B included 33 patients with idiopathic infertility (23 with asthenozoospermia and 10 with oligozoospermia), and Group C (the control group) included 30 healthy fertile men.

Data Collection

Participants underwent a comprehensive assessment, including the following aspects:

History: This included information such as patient age, age of puberty onset, age of varicoccele onset (if applicable), sexual history, number of children, lifestyle habits (smoking, alcohol, drug use), medical history (using cytotoxic, teratogenic, or antiandrogen drugs), surgical history, spinal cord trauma, prostatectomy, sexually transmitted diseases, and epididymitis or epididymo-orchitis.

Examination: General and genital examinations were conducted, encompassing secondary sexual characteristics, body musculature, tall span index, gynecomastia, and body mass index, as well as a thorough examination of the penis, scrotum, epididymis, vas deferens, and spermatic cord.

Scrotal Duplex: Scrotal duplex examinations were performed to identify the presence of varicocele.

Semen Analysis: Semen samples were collected following WHO Manual (2010), with a recommended abstinence period of 2-5 days. Samples were collected by masturbation in sterile containers without the use of lubricants or soap. Samples were incubated at 37°C until complete liquefaction occurred (30-60 minutes). Dynamic and morphological analyses were conducted using CASA (Computer-Assisted Semen Analysis, MIRALAB, ISO9001, ISO13485) to assess sperm parameters.

Exclusion Criteria: Patients with conditions such as erectile dysfunction, benign prostatic hyperplasia, psychological disorders, genetic sex disorders, azoospermia, necrozoospermia, severe debilitating diseases, malnutrition, or use of cytotoxic, teratogenic, or antiandrogen drugs were excluded from the study.

Statistical analysis was performed using the statistical software package SPSS version 22.0 (SPSS Inc, Armonk, NY:

IBM Corp). For the descriptive analysis, results are presented as mean (M) \pm standard deviation (SD). Multiple comparisons were performed with one-way ANOVA and Tukey HSD post-hoc test. Group comparisons with respect to categorical variables are performed using chi-square tests A probability value of *P*<0.05 was considered statistically significant.

Results

The mean age of patients was 31.6 ± 5.81 , 31.3 ± 6.0 , and 29.47 ± 4.27 years in Groups A, B, and C, respectively (*P*>0.05). Primary infertility was diagnosed in 27(71.1%) patients of Group A and 21(63.6%) patients of Group B (Table 1). In Group A, the majority of patients had varicocele grade II (55.3%), followed by grade III (39.5%) and grade I (5.3%) (*P*=0.0006) (Table 2). In Group B, 10(30.3%) patients had oligoasthenozoospermia, and 23(69.7%) patients had asthenozoospermia.

Table 1.

Type of infertility in the patients of the study groups.

Type of infertility	Group A (n= 38)	Group B (n=33)	Statistics
Primary	27 (71.1%)	21 (63.6%)	χ2=0.444 df=1
Secondary	11 (28.9)	12 (14.1%)	P=0.505

Table 2.

Distribution of patients in Group A according to the varicocele grade.

Grade of varicocele	Group A (n=38)	Statistics
Grade I	2 (5.3)	
Grade II	21 (55.3%)	χ2=14.895 df=2 P=0.0006
Grade III	15 (39.5%)	

In terms of semen parameters, the mean semen volume was significantly lower in Group A than in Group C (2.68±0.99 mL vs. 3.31±1.05 mL, P=0.0219); however, mean pH did not show significant differences between study groups, but the mean liquefaction time of semen was slightly higher in Groups A and B than in Group C, without statistical significance (Table 3). We found that sperm concentration, total sperm count, sperm progressive motility, and sperm progressive+non-progressive motility were significantly lower in Group A and Group B than in Group C; however, there were no differences between Group A and Group B regarding these parameters (Table 4). The sperm morphology index was significantly lower in Group A than in Group C (P=0.0024); no differences were found between Group B and Group C and Group B and Group A (Table 5). According to the anatomic-morphological characteristics of sperm, Group A was characterized by significantly smaller dimensions of the length and width of the head, its area, and perimeter, as well as the acrosome coverage, compared to both Group C and Group B (Table 6). The mean value of MAI and TZI did not significantly differ between study groups (P=0.2573 and P=0.2480, respectively). However, the mean value of SDI was significantly lower in Group A than in Group C (P=0.004) (Table 7).

Table 3.

Semen analysis (macroscopic examination) in the study groups.

Parameter	Group A (1)	Group B (2)	Group C (3)	Statistics
Volume, mL	2.68±0.99	3.17±0.81	3.31±1.05	$\begin{array}{c} F=\!$
pH	7.51±0.12	7.52±0.11	7.50± 0.11	F=0.2426 P=0.7851
Liquefaction time, min	27.89±14.73	28.21±11.49	21.73±5.17	$\begin{array}{c} F=3.1638\\ P=0.0466\\ P=0.9924\\ P^{1-2}=0.0783\\ P^{1-3}_{2-3}=0.0715 \end{array}$

Table 4.

Semen analysis (microscopic examination) in the study groups.

Parameter	Group A (1)	Group B (2)	Group C (3)	Statistics
Sperm concentration, 10 ⁶ /ml	23.63±24.97	20.73±24.02	55.88±22.71	$\begin{array}{l} F=\!20.7786\\ P=\!0.0000\\ P_{-2}\!=\!0.8678\\ P_{-3}^{1-2}\!=\!0.0000\\ P_{2-3}^{-3}\!=\!0.0000 \end{array}$
Total sperm count, 10 ⁶ /ml	62.57±76.51	79.81±110.29	178.79±88.66	$\begin{array}{l} F=\!14.8199\\ P=\!0.0000\\ P_{-\!2}\!=\!0.7128\\ P_{1^{-2}}\!=\!0.0000\\ P_{2^{-3}}\!=\!0.0001 \end{array}$
Progressive motility, %	18.39±18.01	14.36±13.44	52.91±12.63	$\begin{array}{l} F{=}61.7070\\ P{=}0.0000\\ P_{-}{=}0.5041\\ P_{1{-}2}{=}0.0000\\ P_{2{-}3}{=}0.0001 \end{array}$
Progressive + non- progressive motility, %	32.18±23.04	26.05±15.57	69.08±13.70	$\begin{array}{l} F{=}50.7048\\ P{=}0.0000\\ P_{=}{=}0.3411\\ P_{1{-}2}{=}0.0000\\ P_{2{-}3}{=}0.0001 \end{array}$

Table 5.

The sperm morphology index in the study groups.

Parameter	Group A (1)	Group B (2)	Group C (3)	Statistics
Sperm morphology index, %	23.18±18.63	28.16±13.40	35.93±11.58	$\begin{array}{c} F{=}5.9615\\ P{=}0.0036\\ P_{1{-}2}{=}0.3543\\ P_{1{-}3}{=}0.0024\\ P_{2{-}3}{=}0.1095 \end{array}$

Table 6.

The anatomic-morphological characteristics of sperm in the study groups.

Parameter	Group A (1)	Group B (2)	Group C (3)	Statistics
Head length, µm	3.60±2.04	4.74±0.23	4.78±0.21	$\begin{array}{c} F=\!9.9490\\ P=\!0.0001\\ P_{-2}\!=\!0.0008\\ P_{-3}^{1-2}\!=\!0.0007\\ P_{2-3}^{2-3}\!=\!0.9913 \end{array}$
Head width, μm	2.27±1.29	2.95±0.15	2.94±0.15	$\begin{array}{c} F=\!8.4143\\ P=\!0.0004\\ P_{-\!\!\!\!\!=0.0016}\\ P_{-\!\!\!\!=0.0026}^{1\cdot3}=\!0.9520 \end{array}$
Length/ width ratio	1.22±0.69	1.62±0.10	1.63±0.09	$\begin{array}{c} F{=}10.4762\\ P{=}0.0001\\ P_{}{=}0.0005\\ P_{}{}^{1{-}2}{=}0.0005\\ P_{}{}^{2{-}3}{=}0.9968 \end{array}$
Head area, μm^2	8.44±4.80	11.00±0.84	11.13±0.70	$\begin{array}{c} F=8.9930\\ P=0.0003\\ P_{-2}=0.0016\\ P_{-3}^{1-2}=0.0012\\ P_{2-3}=0.9839 \end{array}$
Head perimeter, μm	9.81±5.55	12.87±0.55	12.93±0.44	$\begin{array}{c} F=7.5856\\ P=0.0009\\ P_{=0.0009}\\ P_{=0.0254}^{1-3}=0.5975 \end{array}$
Acrosome coverage, %	29.86±19.43	38.97±8.56	33.72±13.69	$\begin{array}{c} F=3.3092\\ P=0.0407\\ P_{-2}=0.0311\\ P_{-3}^{1-3}=0.5405\\ P_{2-3}^{1-3}=0.3464 \end{array}$

Table 7.

Sperm morphology indices in the study groups.

	Group A (1)	Group B (2)	Group C (3)	Statistics
MAI	2.01±0.37	2.11±0.33	1.97±0.34	F=1.3764 P=0.2573
TZI	1.06±0.15	1.05±0.14	1.01 ± 0.06	F=1.4142 P=0.2480
SDI	0.73±0.24	0.68±0.20	0.57±0.13	$\begin{array}{l} F{=}5.5087 \ P{=}0.0054 \\ P_{1{-}2}{=}0.5454 \ P_{1{-}3}{=}0.0040 \\ P_{2{-}3}{=}0.0787 \end{array}$

Discussion

Varicocele is a common condition found in 15% of the general population and 19%-41% of infertile males,^(1-3,21) making it the second most prevalent cause of infertility after idiopathic infertility. Despite the considerable frequency of varicocele in subfertile individuals and proven spermatogenic failure, the specific mechanisms behind varicocele's negative impact on fertility remain unclear.⁽²²⁾ Nevertheless, it affects all sperm characteristics, including count, motility, and morphology.⁽²³⁾

Sperm morphology, a reflection of intricate cellular changes during spermiogenesis, has been identified by some experts as a particularly robust predictor of fertility.^(12,24) This association between sperm morphology and fertility has been

established in numerous species, emphasizing the critical role of sperm morphology in fertility assessment.⁽²⁵⁾ Beyond mere motility, sperm morphology encapsulates vital genetic and DNA characteristics.⁽²⁶⁾

Our investigation uncovered a strong link between infertility and sperm morphology across three distinct groups: healthy fertile males, subfertile individuals with varicocele, and those with idiopathic infertility. Healthy fertile males show normal semen characteristics (volume, count, motility, and morphology), as reported by Aziz et al.⁽²⁷⁾ and Ahmad et al.⁽²⁸⁾ Based on semen characteristics and the existence of a varicocele, subfertile individuals were divided into groups, as previously investigated by Pasqualotto et al.⁽²⁹⁾ and Blumer et al.⁽³⁰⁾

We found that sperm concentration, total sperm count, sperm progressive motility, and sperm progressive+nonprogressive motility were significantly lower in patients with varicocele and idiopathic infertile males than in healthy fertile controls (P=0.000 in all cases). This aligns with findings reported by Vivas-Acevedo et al.,⁽³¹⁾ highlighting decreased sperm motility in infertile males with varicoceles. However, it is worth noting that Saleh and Agarwal⁽³²⁾ found no substantial disparities in sperm motility between infertile males and fertile controls.

Furthermore, our study revealed that the mean sperm morphology index was significantly lower in subfertile patients with varicocele than in healthy fertile men (P=0.0024). These observations align with the results of Tawadrous et al.,⁽³³⁾ Mostafa et al.,⁽³⁴⁾ and Vivas-Acevedo,⁽³⁵⁾ who reported similar findings.

Conversely, the WHO study indicated that infertile males with varicocele exhibited reduced sperm concentration but did not provide specific evidence concerning motility and morphology.⁽³⁶⁾ Some researchers postulate that the observed low sperm concentration may be attributed to the elevated rate of germ apoptosis often found in men. In contrast, diminished motility may be linked to a high concentration of reactive oxygen species or anti-sperm antibodies.^(37,38)

Semen analysis normally evaluates only the dimensions of the sperm head (WHO, 1999)⁽¹²⁾ because head morphological anomalies significantly affect male fertility.⁽³⁹⁾ However, despite WHO recommendations to consider additional aspects of sperm morphology, little attention has been given to the diameters of the midpiece and flagellum.⁽⁴⁰⁾

Our study revealed significant deviations in head lengths, perimeters, and acrosome coverage in patients of the studied groups. Subfertile patients with varicocele were characterized by significantly smaller dimensions of the length and width of the head, its area, and perimeter, as well as the acrosome coverage than in fertile men.

These findings echo the results of Vazquez Levin⁽⁴¹⁾ and Schatte,⁽⁴²⁾ who identified a lower frequency of morphologically normal forms in varicocele patients when stringent criteria were applied. In contrast, Saleh and Agarwal⁽³²⁾ observed no significant differences in sperm morphology between infertile individuals and fertile controls. MacLeod in 1965⁽⁴³⁾ identified the "stress pattern," characterized by elongated tapering sperm heads and amorphous spermatozoa linked with varicocele. However, Rodrigues-Rigau et al.⁽⁴⁴⁾ found no notable changes in sperm shape between males with and without varicocele. WHO (1999) also observed a substantial negative correlation between average head length and the proportion of sperm with "normal" morphology.⁽¹²⁾

Our study also revealed a significantly reduced sperm deformity index in subfertile patients with varicocele, compared to fertile men, suggesting that increased abnormality in head length and perimeters is one of the possible causes of infertility due to varicocele. Wang et al.(45) observed that a 1°C increase in testicular temperature inhibits spermatogenesis by 14%, resulting in a drop in sperm production. Additionally, exposure to extreme temperatures alters the shape of sperm, resulting in a rise in sperm with aberrant morphology. Within 6-8 months of exposure to high temperatures, the average percentage of sperm with aberrant morphology increases from 30% to 60%. The researchers hypothesized that heating the testes decreased the quantity and the quality of sperm production.⁽⁴⁵⁾ Activation of the p53 gene, a tumor-suppressor gene expressed in testes, is a well-known mechanism for explaining spermatogenic dysfunction caused by heat.^(46,47) It is most highly expressed in pachytene spermatocytes.⁽⁴⁸⁾ High scrotal temperatures result in condensation of nuclear chromatin, which activates p53 and halts the cell cycle. This hinders the clonal expansion of germ cells with DNA damage. Morgentaler et al.(49) hypothesized that p53 may be involved in heat-induced germ-cell death. p53 is situated on the nuclear membrane of normal germ cells and is responsible for germ-cell quality control. With heat-induced nuclear damage, it translocates to the nucleoplasm and triggers germ-cell death.(50)

In conclusion, our study highlights the significant association between sperm morphology and male infertility in varicocele and idiopathic subfertile males. Further research is needed to explore the relationship between sperm morphometry, sperm function, and fertility across different species. Additionally, understanding the therapeutic implications of sperm morphology could aid in selecting semen samples with the least aberrant morphometry for subfertile men.

Competing Interests

The authors declare that they have no competing interests.

Acknowledgments

This study was funded by the Prince Sattam bin Abdulaziz University (PSAU/2023/R/1444) and Princess Nourah bint Abdulrahman University Researchers Supporting (PNURSP2023R99), Riyadh, Saudi Arabia.

References

1. Zegers-Hochschild F, Adamson GD, de Mouzon J, Ishihara O, Mansour R, Nygren K, Sullivan E, van der Poel S; International Committee for Monitoring Assisted Reproductive Technology; World Health Organization. The International Committee for Monitoring Assisted Reproductive Technology (ICMART) and the World Health Organization (WHO) Revised Glossary on ART Terminology, 2009. Hum Reprod. 2009 Nov;24(11):2683-7. doi: 10.1093/humrep/dep343.

2. Speroff L, Glass RH, Kase NG. Clinical Gynecologic Endocrinology and Infertility. 6th Edition, Lippincott Williams & Wilkins, Philadelphia; 1999.

3. Lotti F, Maggi M. Sexual dysfunction and male infertility. Nat Rev Urol. 2018 May;15(5):287-307.

4. Skoracka K, Eder P, Łykowska-Szuber L, Dobrowolska A, Krela-Kaźmierczak I. Diet and Nutritional Factors in Male (In)fertility-Underestimated Factors. J Clin Med. 2020 May 9;9(5):1400. doi: 10.3390/jcm9051400.

5. Zhang X, Zhang J, Cai Z, Wang X, Lu W, Li H. Effect of unilateral testicular torsion at different ages on male fertility. J Int Med Res. 2020 Apr;48(4):300060520918792. doi: 10.1177/0300060520918792.

6. Hallast P, Kibena L, Punab M, Arciero E, Rootsi S, Grigorova M, et al. A common 1.6 mb Y-chromosomal inversion predisposes to subsequent deletions and severe spermatogenic failure in humans. Elife. 2021 Mar 30;10:e65420. doi: 10.7554/ eLife.65420.

7. Turner KA, Rambhatla A, Schon S, Agarwal A, Krawetz SA, Dupree JM, Avidor-Reiss T. Male Infertility is a Women's Health Issue-Research and Clinical Evaluation of Male Infertility Is Needed. Cells. 2020 Apr 16;9(4):990. doi: 10.3390/ cells9040990.

8. Gorelick JI, Goldstein M. Loss of fertility in men with varicocele. Fertil Steril. 1993 Mar;59(3):613-6.

9. Naughton CK, Nangia AK, Agarwal A. Varicocele and male infertility: P:art II - Pathophysiology of varicocele in male infertility. Hum Reprod Update. 2001;7(5):473-481.

10. Chan CC, Sun GH, Shui HA, Wu GJ. Differential spermatozoal protein expression profiles in men with varicocele compared to control subjects: upregulation of heat shock proteins 70 and 90 in varicocele. Urology. 2013 Jun;81(6):1379. e1-8. doi: 10.1016/j.urology.2013.01.031.

11. Samplaski MK, Agarwal A, Sharma R, Sabanegh E. New generation of diagnostic tests for infertility: review of specialized semen tests. Int J Urol. 2010 Oct;17(10):839-47. doi: 10.1111/j.1442-2042.2010.02619.x. Erratum in: Int J Urol. 2011 Mar;18(3):262.

12. World Health Organization. WHO Laboratory Manual for the examination of human semen and semen-cervical mucus interaction. Fourth edition. Cambridge, UK: Cambridge University Press; 1999.

13. Kruger TF, Lacquet FA, Sarmiento CA, Menkveld R, Ozgür K, Lombard CJ, Franken DR. A prospective study on the predictive value of normal sperm morphology as evaluated by computer (IVOS). Fertil Steril. 1996 Aug;66(2):285-91. doi: 10.1016/s0015-0282(16)58455-6.

14. Hidalgo M, Rodríguez I, Dorado J. Influence of staining and sampling procedures on goat sperm morphometry using the Sperm Class Analyzer. Theriogenology. 2006 Sep 1;66(4):996-1003. doi: 10.1016/j.theriogenology.2006.02.039.

15. Filimberti E, Degl'Innocenti S, Borsotti M, Quercioli M, Piomboni P, Natali I, et al. High variability in results of semen analysis in andrology laboratories in Tuscany (Italy): the experience of an external quality control (EQC) programme. Andrology. 2013 May;1(3):401-7.

16. Verstegen J, Iguer-Ouada M, Onclin K. Computer assisted semen analyzers in andrology research and veterinary practice. Theriogenology. 2002 Jan 1;57(1):149-79. doi: 10.1016/s0093-691x(01)00664-1.

17. Yániz JL, Capistrós S, Vicente-Fiel S, Soler C, Nuñez de Murga J, Santolaria P. Study of nuclear and acrosomal

sperm morphometry in ram using a computer-assisted sperm morphometry analysis fluorescence (CASMA-F) method. Theriogenology. 2014 Oct 1;82(6):921-4.

18. Yániz JL, Vicente-Fiel S, Capistrós S, Palacín I, Santolaria P. Automatic evaluation of ram sperm morphometry. Theriogenology. 2012 Apr 15;77(7):1343-50. doi: 10.1016/j. theriogenology.2011.10.039.

19. Soler C, García-Molina A, Sancho M, Contell J, Núñez M, Cooper TG. A new technique for analysis of human sperm morphology in unstained cells from raw semen. Reprod Fertil Dev. 2016 Mar;28(4):428-33. doi: 10.1071/RD14087.

20. Rrumbullaku L, Boci R, Dedja A, Dautaj K. Sperm morphology in infertile men with varicocele. 1st Balkan Symposium of Andrology. Alexandroupolis, Greece; June 12-14, 1998.

21. Mancini A, Festa R, Raimondo S, Silvestrini A, Giacchi E, Littarru GP, Pontecorvi A, Meucci E. Biochemical alterations in semen of varicocele patients: a review of the literature. Adv Urol. 2012;2012:903931. doi: 10.1155/2012/903931.

22. Saleh RA, Agarwal A, Sharma RK, Said TM, Sikka SC, Thomas AJ Jr. Evaluation of nuclear DNA damage in spermatozoa from infertile men with varicocele. Fertil Steril. 2003 Dec;80(6):1431-6. doi: 10.1016/s0015-0282(03)02211-8.

23. Al-AliBM, Marszalek M, Shamloul R, Pummer K, Trummer H. Clinical parameters and semen analysis in 716 Austrian patients with varicocele. Urology. 2010 May;75(5):1069-73. doi: 10.1016/j.urology.2009.11.042.

24. Nallella KP, Sharma RK, Aziz N, Agarwal A. Significance of sperm characteristics in the evaluation of male infertility. Fertil Steril. 2006 Mar;85(3):629-34.

25. Al-Makhzoomi A, Lundeheim N, Håård M, Rodríguez-Martínez H. Sperm morphology and fertility of progenytested AI dairy bulls in Sweden. Theriogenology. 2008 Sep 1;70(4):682-91. doi: 10.1016/j.theriogenology.2008.04.049.

26. Murphy C, Fahey AG, Shafat A, Fair S. Reducing sperm concentration is critical to limiting the oxidative stress challenge in liquid bull semen. J Dairy Sci. 2013 Jul;96(7):4447-54.

27. Aziz N, Agarwal A, Nallella KP, Thomas AJ Jr. Relationship between epidemiological features and aetiology of male infertility as diagnosed by a comprehensive infertility service provider. Reprod Biomed Online. 2006 Feb;12(2):209-14.

28. Ahmad L, Jalali S, Shami SA, Akram Z. Sperm preparation: DNA damage by comet assay in normo- and teratozoospermics. Arch Androl. 2007 Nov-Dec;53(6):325-38.

29. Pasqualotto FF, Sharma RK, Pasqualotto EB, Agarwal A. Poor semen quality and ROS-TAC scores in patients with idiopathic infertility. Urol Int. 2008;81(3):263-70.

30. Blumer CG, Fariello RM, Restelli AE, Spaine DM, Bertolla RP, Cedenho AP. Sperm nuclear DNA fragmentation and mitochondrial activity in men with varicocele. Fertil Steril. 2008 Nov;90(5):1716-22. doi: 10.1016/j.fertnstert.2007.09.007.

31. Vivas-Acevedo G, Lozano JR, Camejo MI. Effect of varicocele grade and age on seminal parameters. Urol Int. 2010;85(2):194-9. doi: 10.1159/000314226.

32. Saleh RA, Agarwal A. Oxidative stress and male infertility: from research bench to clinical practice. J Androl. 2002 Nov-Dec;23(6):737-52.

33. Tawadrous GA, Aziz AA, Mostafa T. Seminal soluble fas relationship with oxidative stress in infertile men with varicocele. Urology. 2013 Oct;82(4):820-3.

34. Mostafa T, Rashed L, Nabil N, Amin R. Seminal BAX and BCL2 gene and protein expressions in infertile men with varicocele. Urology. 2014 Sep;84(3):590-5. doi: 10.1016/j.

urology.2014.05.016.

35. Vivas-Acevedo G, Lozano-Hernández R, Camejo MI. Varicocele decreases epididymal neutral α -glucosidase and is associated with alteration of nuclear DNA and plasma membrane in spermatozoa. BJU Int. 2014 Apr;113(4):642-9. doi: 10.1111/bju.12523.

36. The influence of varicocele on parameters of fertility in a large group of men presenting to infertility clinics. World Health Organization. Fertil Steril. 1992 Jun;57(6):1289-93.

37. Marmar JL. The pathophysiology of varicoceles in the light of current molecular and genetic information. Hum Reprod Update. 2001 Sep-Oct;7(5):461-72. doi: 10.1093/ humupd/7.5.461.

38. Agarwal A, Virk G, Ong C, du Plessis SS. Effect of oxidative stress on male reproduction. World J Mens Health. 2014 Apr;32(1):1-17. doi: 10.5534/wjmh.2014.32.1.1.

39. Barratt CL, Mansell S, Beaton C, Tardif S, Oxenham SK. Diagnostic tools in male infertility-the question of sperm dysfunction. Asian J Androl. 2011 Jan;13(1):53-8. doi: 10.1038/aja.2010.63.

40. Aziz N, Buchan I, Taylor C, Kingsland CR, Lewis-Jones I. The sperm deformity index: a reliable predictor of the outcome of oocyte fertilization in vitro. Fertil Steril. 1996 Dec;66(6):1000-8. doi: 10.1016/s0015-0282(16)58697-x.

41. Vazquez-Levin MH, Friedmann P, Goldberg SI, Medley NE, Nagler HM. Response of routine semen analysis and critical assessment of sperm morphology by Kruger classification to therapeutic varicocelectomy. J Urol. 1997 Nov;158(5):1804-7. doi: 10.1016/s0022-5347(01)64134-x.

42. Schatte EC, Hirshberg SJ, Fallick ML, Lipschultz LI, Kim ED. Varicocelectomy improves sperm strict morphology and motility. J Urol. 1998 Oct;160(4):1338-40.

43. MacLeod J. Seminal cytology in the presence of varicocele. Fertil Steril. 1965 Nov-Dec;16(6):735-57. doi: 10.1016/s0015-0282(16)35765-x.

44. Rodriguez-Rigau LJ, Smith KD, Steinberger E. Varicocele and the morphology of spermatozoa. Fertil Steril. 1981 Jan;35(1):54-7. doi: 10.1016/s0015-0282(16)45258-1.

45. Wang C, McDonald V, Leung A, Superlano L, Berman N, Hull L, Swerdloff RS. Effect of increased scrotal temperature on sperm production in normal men. Fertil Steril. 1997 Aug;68(2):334-9. doi: 10.1016/s0015-0282(97)81525-7.

46. Rogel A, Popliker M, Webb CG, Oren M. p53 cellular tumor antigen: analysis of mRNA levels in normal adult tissues, embryos, and tumors. Mol Cell Biol. 1985 Oct;5(10):2851-5. doi: 10.1128/mcb.5.10.2851-2855.1985.

47. Almon E, Goldfinger N, Kapon A, Schwartz D, Levine AJ, Rotter V. Testicular tissue-specific expression of the p53 suppressor gene. Dev Biol. 1993 Mar;156(1):107-16.

48. Schwartz D, Goldfinger N, Rotter V. Expression of p53 protein in spermatogenesis is confined to the tetraploid pachytene primary spermatocytes. Oncogene. 1993 Jun;8(6):1487-94.

49. Morgentaler A, Stahl BC, Yin Y. Testis and temperature: an historical, clinical, and research perspective. J Androl. 1999 Mar-Apr;20(2):189-95.

50. Yin Y, DeWolf WC, Morgentaler A. p53 is associated with the nuclear envelope in mouse testis. Biochem Biophys Res Commun. 1997 Jun 27;235(3):689-94.

*Corresponding author: Ramadan S. Hussein. Department of Internal Medicine, College of Medicine, Prince Sattam Bin Abdulaziz University, Al-Kharj, Saudi Arabia. E-mail: ramadangazeera@ yahoo.com